

Associated Institute of





DEVELOPMENT OF AN ANALYSIS METHOD TO ASSESS THE OCCUPATIONAL RISK DEALING WITH THERAPEUTIC MONOCLONAL ANTIBODIES USING LIQUID CHROMATOGRAPHY AND HIGH-RESOLUTION MASS SPECTROMETRY (LC-HRMS)

Lars M. H. Reinders^{1,2}, Martin D. Klassen¹, Martin Jaeger², Thorsten Teutenberg¹, Jochen Tuerk¹

Institut f
ür Energie- und Umwelttechnik e. V. (IUTA, Institute of Energy and Environmental Technology), Bliersheimer Str. 58-60, 47229 Duisburg, Germany, teutenberg@iuta.de
 Hochschule Niederrhein (University of Applied Science Niederrhein), Reinarzstr. 49, 47805 Krefeld, Germany

Introduction

Conclusion and Outlook

- In an ongoing debate about the health risk assessment of monoclonal antibodies (mAbs) the following points need to be considered:
 - Long-term low dose data is not available
 - Pulmonary uptake of lyophilized powder or mAbs in aerosols is the most probable route of entry
 - Risk of sensitization outgoing from long-term low doses cannot be excluded
 - Safety recommendation varies greatly, from protective gloves alone to full protective clothing
- We present the development of an analytical method that enables us to determine the occupational exposure
 - o Intact mAbs, heavy chain, light chain or peptides were investigated as suitable starting points
 - Analysis of intact or reduced mAbs leads to a large, sensitivity decreasing charge pattern
- Development of a peptide based analytical workflow to quantify monoclonal antibodies (mAbs) as single substances as well as sum parameter is shown
- Limit of detections between 1 µg/sample for daratumumab and 25 µg/sample for the sum parameter were reached
- Stability of peptides in 24 h long-term air sampling was proved
- Recovery rates between 80% and 120% were observed
- Future investigations will be performed to collect experimental data of the occupational exposure of pharmacy staff to
- Peptide based workflows need extensive sample preparation, which can induce chemical modifications
- Peptide level offers the opportunity to use a sum parameter for all mAbs
- \circ Expected occupational exposition is in the range of several µg/working shift

airborne mAbs

 The resulting data will help to build a solid base for further discussions about the occupational health

Analytical Workflow

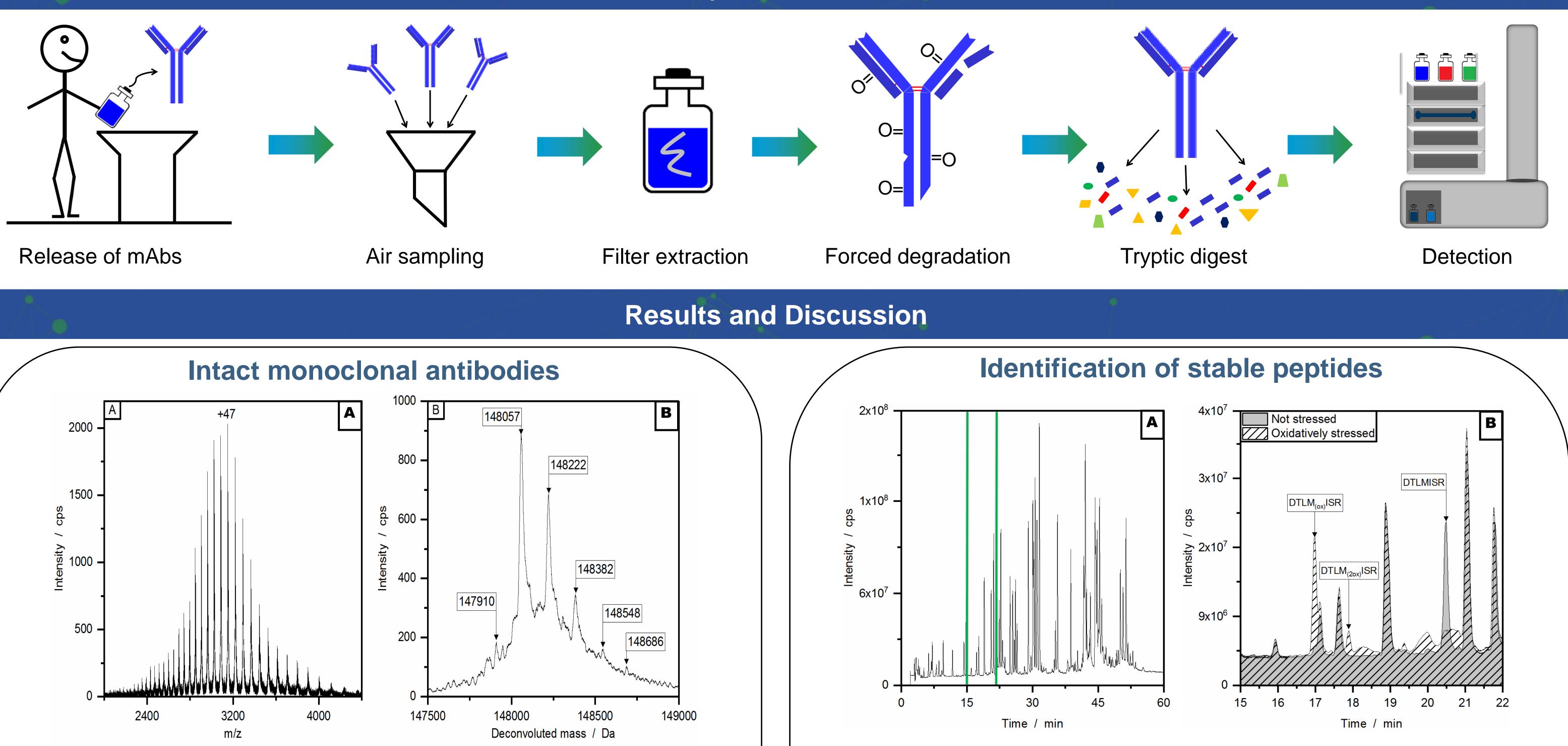


Figure 1: A: Mass-to-charge distribution of trastuzumab at denaturizing conditions by LC-ESI-MS with z = +47 as most intensive signal. B: The deconvolved masses are about 148 kDa depending on their glycosylation profile.

- LC-ESI-MS of monoclonal antibodies leads to a large charge pattern
 - Calculation of the MW with a deconvolving algorithm such as MaxEnt
- Intensity distribution over the single charge states leads to a sensitivity loss
 Peptide level offers higher sensitivity, due to less charge distribution

Air sampling

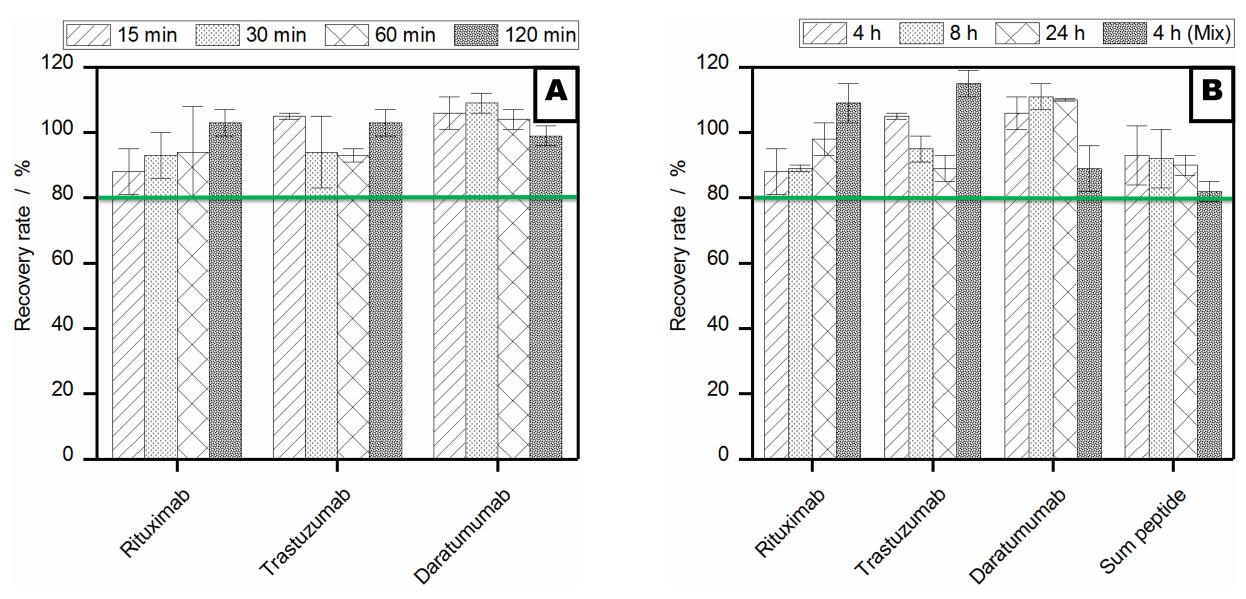


Figure 2: A: Chromatogram of a tryptic digest of trastuzumab. B: Enlargement from 15 min to 22 min. The effects of the oxidising agent H_2O_2 (lined) on trastuzumab are compared with an NaCl solution (grey) treated sample. The H_2O_2 treated sample was incubated. The NaCl solution was adjusted to physiological conditions.

- Tryptic digests of monoclonal antibodies lead to a large number of peptides
 - Only one signature peptide for the single substances and one collective peptide to create a sum parameter is necessary
- Instable peptides are not suitable for an analytical workflow
 - Forced degradation tests with an acidic, a basic and an oxidative medium
- Stable and chosen peptides

0	Rituximab	FSGSGSGTSYSLTISR	(<i>m/z</i> 803.8890; z = 2)
0	Trastuzumab	LLIYSASFLYSGVPSR	(<i>m/z</i> 886.9827; z = 2)
0	Daratumumab	LLIYDASNR	(<i>m/z</i> 532.7904; z = 2)
0	Sum parameter	DSTYSLSSTLTLSK	(<i>m/z</i> 751.8828; z = 2)

Figure 3: A: PVDF-filter were spiked with monoclonal antibodies solutions and extracted using pH 7.4 phosphate buffered saline (PBS) at 30 ° C and varying extraction time. B: The spiked PVDF-filter were connected to a membrane pump with a constant airflow of 2 L/min and varying sampling time. The extraction time was constant at 15 min.

- Extraction times of 15 min is sufficient
- Long-term sampling up to 24 h is possible
- No negative interaction between the monoclonal antibodies in a mix

Method validation

Table 1: The LOD and LOQ are defined as S/N = 3 and S/N = 10. Reproducibility was determined on the same day and repeatability on three different days. Samples were analyzed as triplicates. The linearity was determined from the LOQ to 15 and 45 mg/sample, respectively. The test of linearity was performed according to Mandel with a statistical safety of 99.9%.

Peptides for	LOD (µg/sample)	LOQ (µg/sample)	Reproducibility (%)	Repeatability (%)	Linearity (mg/sample)
Rituximab	9.6	33	88 ± 7	92 ± 6	15
Trastuzumab	4.4	15	105 ± 1	97 ± 7	15
Daratumumab	1.4	4.7	106 ± 5	109 ± 4	15
Sum parameter	25	79	97 ± 0.8	95 ± 4	45

Acknowledgement



We thank Agilent Technologies and especially Dr. Bita Kolahgar for providing the HPLC-QTOF system and the technical support.



Abstract number 5PSQ-057 ATC code L01 – Cytostatics