



MIC directed therapy: why, when and what are the pitfalls?

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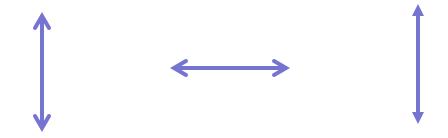
None



Antimicrobial therapy in general



Efficacy of the drug



Potency of a drug (MIC)

Exposure to the bug

In vivo

(PK)



PK/PD indices



- The MIC in included in the indices:
 - %*f*T>MIC
 - AUC/MIC



The reference method











2003 20 june DIN Berlin CEN TC140/WG10

2004 22 april DIN Berlin Combined meeting with ISO ISO/TC 212 WG4 Vienna Agreement

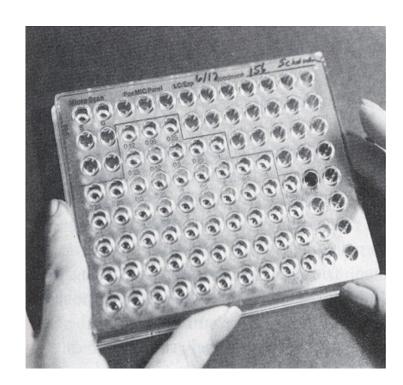
2005 Vote on first draft and comments by all Member Countries

2006 Final version 27 October 2006, 8th CEN, 6th ISO meeting ISO 20776-1

2007 Final version validation ISO 20776-2.







The reference method: microdilution

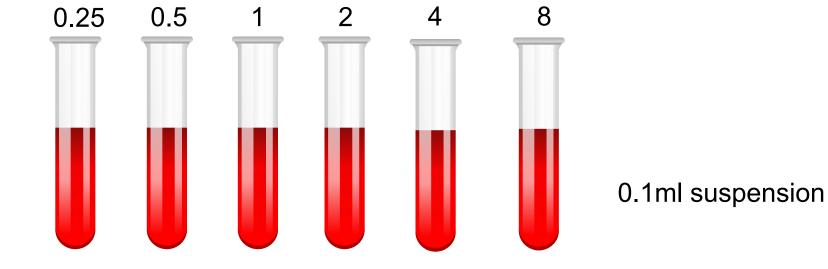
Measure of potency of the antibacterial effect





2-fold increasing antibiotic concentrations in mg/L in Mueller Hinton



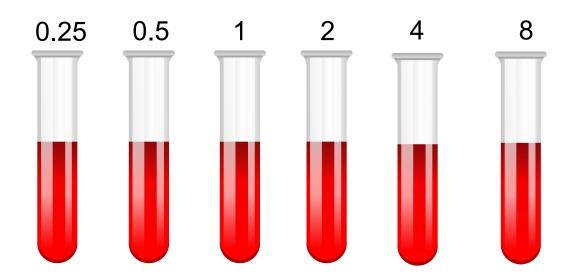


Bacterial suspension: Inoculum 5 (2-8) .10E5 cfu/ml





2-fold increasing antibiotic concentrations in mg/L in Mueller Hinton Bacterial suspension: Inoculum 5 (2-8) .10E5 cfu/ml

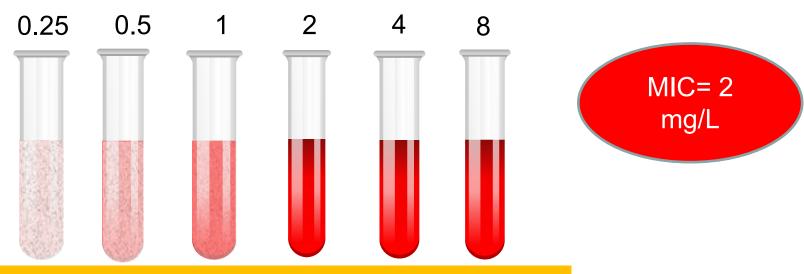


Incubate 36 +/- 1° C 18 +/- 2 hours





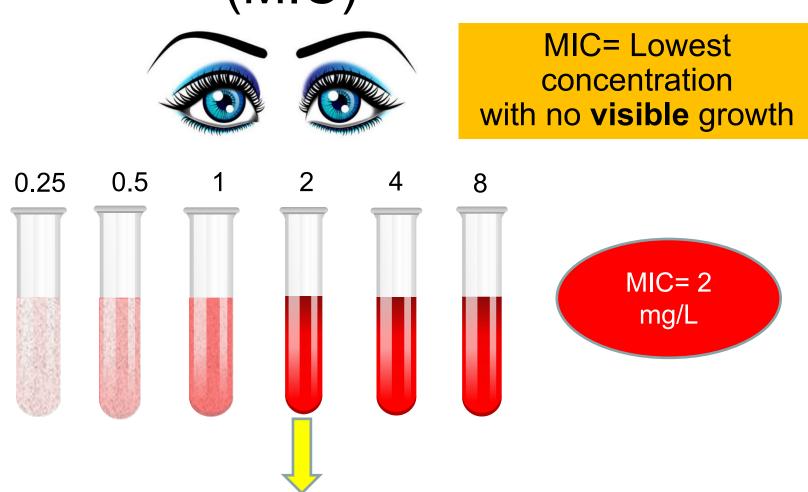
2-fold increasing antibiotic concentrations in mg/L in Mueller Hinton Bacterial suspension: Inoculum 5 (2-8) .10E5 cfu/ml



After inbubation: MIC= Lowest concentration with no **visible** growth







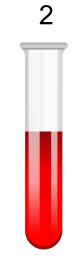
Number of bacteria in this tube varies between 0-10^8 CFU/ml





What happened in the tubes?

Bacterial growth
Bacterial kill
Continuous process over time



Kill



 $\frac{dN}{dt} = \left\{ \lambda \bullet (1 - \frac{N}{N_{\text{max}}}) - \varepsilon \bullet \frac{C^{\gamma}}{C^{\gamma} + EC_{50}^{\gamma}} \right\} \bullet N$

Growth

MIC is the result of these processes over time

Growth rate

Max kill rate

AEM Brussel 20-09-2019 Mouton et al 1997





"The" MIC

Does NOT Quantify bacterial growth

Does NOT Quantify bacterial kill

It is the result of these biological processes over time

High variability and is not very reproducible



The use of other methods



 All methods need to be validated versus the reference method

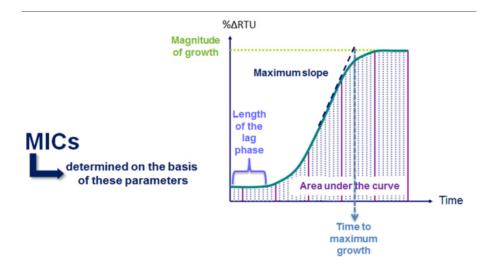






Vitek 2 system, BioMérieux

- algoritm based on a few measurements.
- Not 2-fold dilution.
- Totally different approach
- repetitive turbidimetric monitoring of bacterial growth during an abbreviated incubation period.



Magnitude of growth = Maximum percent change of the transmittance signal (RTU)

Maximum slope = maximum rate of growth during exponential growth phase







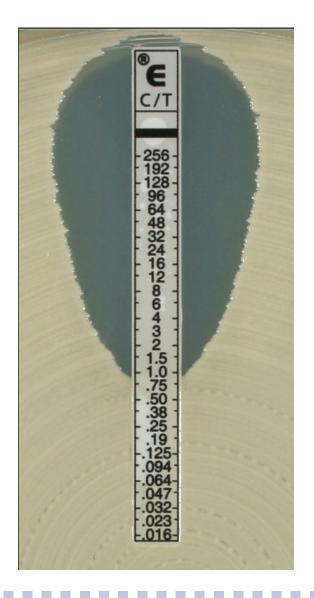
Phoenix system, BD

- * 2-fold dilution,
- * micro-dilution,
- * growth or no-growth (turbidometric and colorimetric (oxidation-reduction indicator) growth detection).
- * different inoculum from the reference method



Gradiënt-tests





- Increasing concentration on the strip
- Antibiotic diffuses into the agar

Ceftolozane/Tazobactam (C/T 256)

Figure: BioMérieux website



Disc diffusion



Do not result in a value in mg/L, but in mm of the zone.



Figure: Thermo Fisher Scientific



Variation in measurements



- Biological variation within one strain
- between-strain variation
- between-laboratory variation
 - Materials
 - people

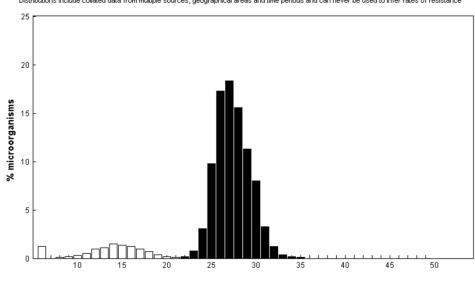


Disc diffusion S aureus and cefoxitin



Cefoxitin / Staphylococcus aureus International wild type zone diameter distribution - Reference database 2019-04-10 EUCAST disk diffusion method

Distributions include collated data from multiple sources, geographical areas and time periods and can never be used to infer rates of resistance



Zone diameter (mm)

Disk content: 30
Epidemiological cut-off (ECOFF): 22 mm (MIC = 4 mg/L)
Wildtype (WT) organisms: ≥ 22 mm (MIC = 4 mg/L)

26134 observations (13 data sources)

~26000 observations (strains) 13 sources (different labs) Different time periods Susceptible strains: 22-35mm



Variation in measurements



- Biological variation within one strain
- between-strain variation
- between-laboratory variation

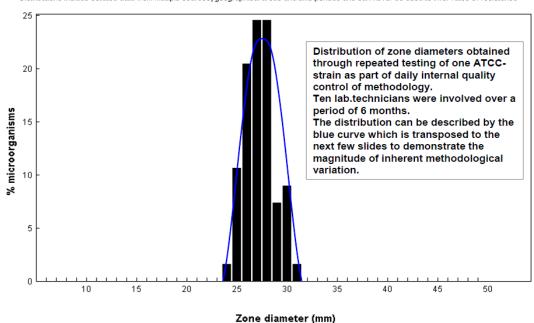


S aureus ATCC 29213 in 1 laboratory



Cefoxitin / Staphylococcus aureus ATCC 29213 EUCAST zone diameter distribution - Reference database EUCAST disk diffusion method

Distributions include collated data from multiple sources, geographical areas and time periods and can never be used to infer rates of resistance



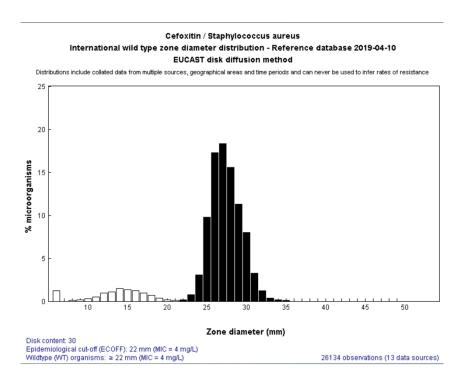
Disk content: 30
Epidemiological cut-off: WT ≥ 22 mm (MIC: -)

122 observations Clinical breakpoints: S ≥ 22 mm, R < 22 mm 1 strain122 measurements6 months10 labtechniciansZones between 24-31 mm



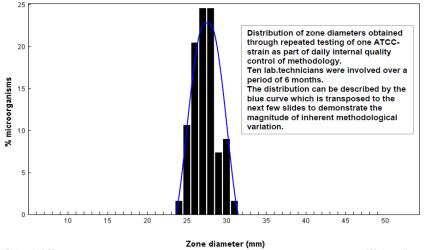
Compare the two distributions





Cefoxitin / Staphylococcus aureus ATCC 29213 EUCAST zone diameter distribution - Reference database EUCAST disk diffusion method

Distributions include collated data from multiple sources, geographical areas and time periods and can never be used to infer rates of resistance



Disk content: 30 Epidemiological cut-off: WT ≥ 22 mm (MIC: -)

Clinical breakpoints: S ≥ 22 mm, R < 22 mm

~26000 strains

1 strain 122 measurements





Second example on the variability

S aureus and linezolid MIC determined by gradienttest (Etest®)

Analysis:

22 different strains

5 different laboratories

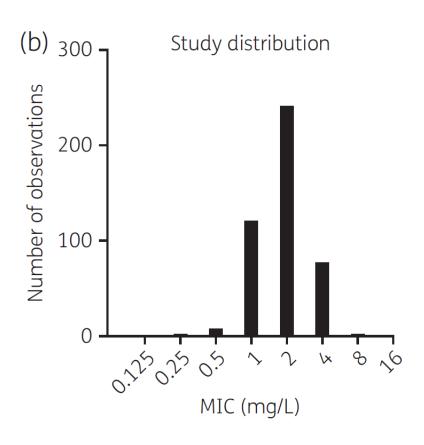
Sent in quadruplicate (blind fashion)

440 observations



MIC-distribution of the strains





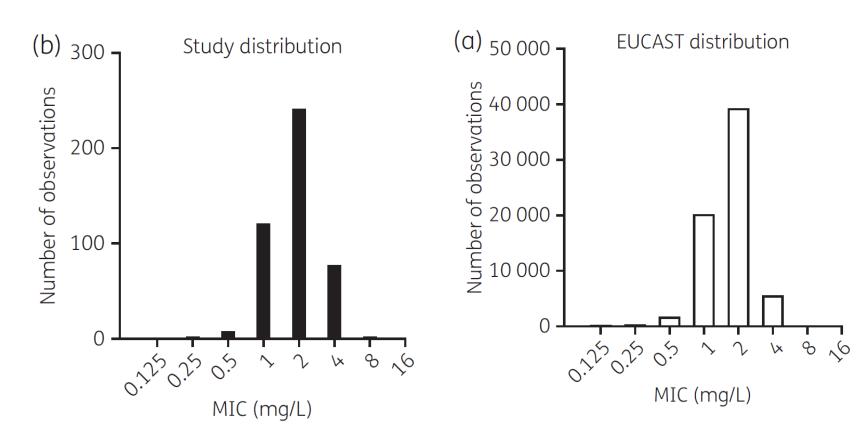
440 observations in 5 labs

→ 20 observations per strain



MIC-distribution of the strains





440 observations in 5 labs

→ 20 observations per strain



Linezolid and S. aureus



Source of the variation

		Sum	of squares (% of to	otal error)			
	Observations (n)	total variation	strain variation	laboratory variation	Explained	R^2	Unexplained assay variation
Total	440	227.82 (100%)	109 22 (47 9%)	23 57 (10 3%)	132 79 (58 3%)	0.58	95.03 (41.7%)

So, "the" MIC does not exists in the routine lab

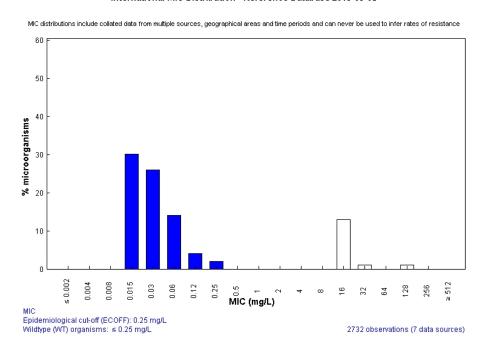


Wild-type distribution

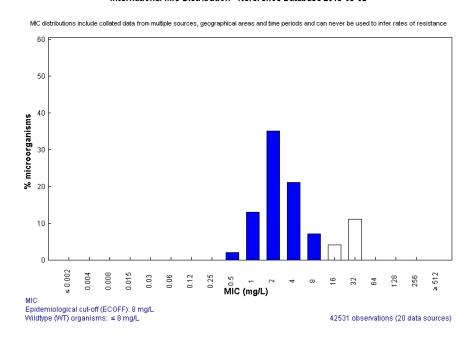


- Distribution of MICs for micro-organisms without resistance mechanisms
- www.eucast.org

Ceftobiprole / Klebsiella pneumoniae International MIC Distribution - Reference Database 2019-09-08



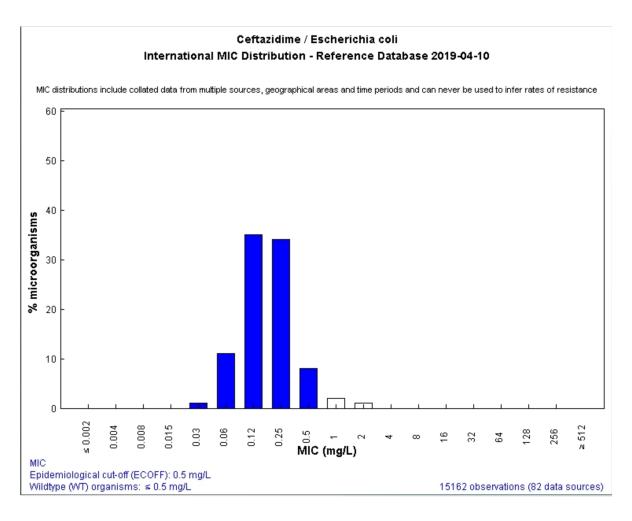
Cefuroxime / Klebsiella pneumoniae International MIC Distribution - Reference Database 2019-09-08





Epidemiological cut-off (ECOFF)





ECOFF: 0.5 mg/L

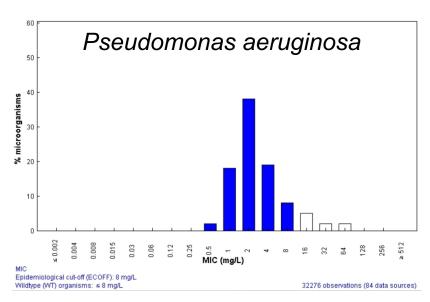


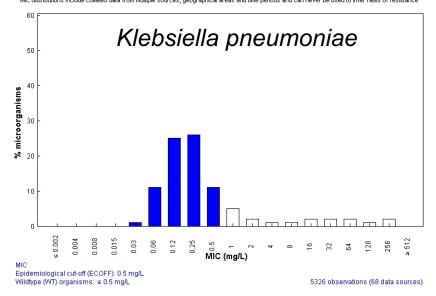


Ceftazidime and *Pseudomonas aeruginosa* and *Klebsiella pneumoniae*

• MIC

<u> </u>	0.002	0.004	0.008	0.016	0.032	0.064	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	ECOFF
Klebsiella pneumoniae	0	0	10	9	89	592	1346	1425	611	281	145	88	104	113	146	136	92	112	27	0.5
<u>Klebsiella spp</u>	0	0	0	15	125	343	351	158	71	43	13	3	3	0	2	0	1	0	0	0.5
Moraxella catarrhalis	0	0	0	6	27	28	10	7	1	1	0	0	0	0	0	0	0	0	0	ND
<u>Morganella morganii</u>	0	2	5	18	67	64	48	38	34	15	12	10	7	19	4	3	0	1	0	0.25
<u>Neisseria gonorrhoeae</u>	0	2	3	12	16	5	0	0	0	0	0	0	0	0	0	0	0	0	0	ND
<u>Proteus mirabilis</u>	0	0	6	86	517	461	92	50	37	30	10	9	2	4	2	2	0	3	0	0.125
<u>Proteus vulgaris</u>	0	0	0	10	52	82	12	2	6	1	1	1	1	1	0	0	0	0	0	0.125
<u>Providencia spp</u>	0	0	0	0	1	9	28	43	6	2	0	4	0	0	0	0	0	0	0	0.5
<u>Providencia stuartii</u>	0	0	0	0	1	1	1	6	4	2	8	6	2	3	1	1	0	1	0	0.5
Pseudomonas aeruginosa	0	0	0	1	4	8	31				12322						167		106	8.0





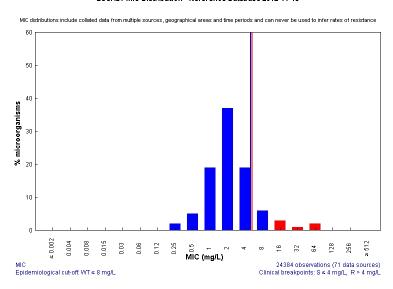


Wild-type distributions



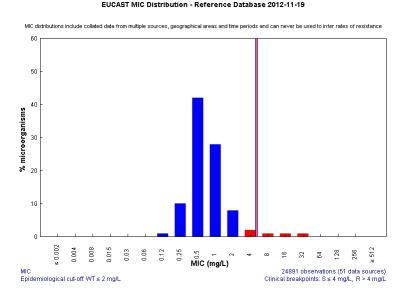
Gentamicin

Gentamicin / Pseudomonas aeruginosa EUCAST MIC Distribution - Reference Database 2012-11-19



Tobramycin

Tobramycin / Pseudomonas aeruginosa



ECOFF 8 mg/L

ECOFF 2 mg/L

Pharmacokinetic profile is very similar PK/PD targets similar



Dosing based on an individual MIC?



NO!



Literature..... example



Int J Antimicrob Agents. 2019 Mar 6. pii: S0924-8579(19)30054-8. doi: 10.1016/j.ijantimicag.2019.03.002. [Epub ahead of print]

Be careful about MICs to amoxicillin for patients with Streptococci-related infective endocarditis.

Pilmis B¹, Lourtet-Hascoët J², Barraud O³, Piau C⁴, Isnard C⁵, Hery-Arnaud G⁶, Amara M⁷, Merens A⁸, Farfour E⁹, Thomas E¹⁰, Jacquier H¹¹, Zahar JR¹², Bonnet E¹³, Le Monnier A², Cattoir V⁴, Corvec S¹⁰, Boutoille D¹⁴, Péan de Ponfilly G¹¹, Reissier S⁵; GMC Study Group.

In multivariate analysis, the only factor associated with in-hospital mortality was MIC for amoxicillin between 0.25 and 2mg/L (p=0.04; OR= 2.23 [1.03-4.88]) whereas protective factor was performance of cardiac surgery for IE (p=0.001, OR = 0.23 [0.1-0.56]).

Population level, not individual level





Conclusion MIC determination and dosing

- Considerable amount of variation
- The only conclusion that can be drawn is, whether the bacteria is within the wild-type distribution or not.
- Do not use such values in individual patient care

Table 1. Suggested interpretation of the MIC for target attainment under various conditions

MIC found	Interpretation for target attainment
Within WT, ≤ECOFF	ECOFF
>ECOFF	MIC + two 2-fold dilutions ^a

^aNumber of dilutions could be higher or lower than two depending on the proficiency of the lab and the drug-species distribution.



Example TDM for sepsis with Klebsiella pneumoniae



- Local lab results reports S for ceftazidime
 - Hopefully the lab does not report a value, such as 0.125mg/L
 - You know that the MIC is maximum 0.5 mg/L

,	0.002	0.004	0.008	0.016	0.032	0.064	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512	ECOFF
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Moraxella catarrhalis	0	0	0	6	27	28	10	7	1	1	0	0	0	0	0	0	0	0	0	ND
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Proteus vulgaris	0	0	0	10	52	82	12	2	6	1	1	1	1	1	0	0	0	0	0	0.125
<u>Providencia spp</u>	0	0	0	0	1	9	28	43	6	2	0	4	0	0	0	0	0	0	0	0.5
Providencia stuartii	0	0	0	0	1	1	1	6	4	2	8	6	2	3	1	1	0	1	0	0.5
Pseudomonas aeruginosa	0	0	0	1	4	8	31	292	966	5975	12322	6271	2738	1712	815	751	167	117	106	8.0

Target value for TDM will be MIC of 0.5 mg/L